



TECH DATA SHEET

REPORTER VIRUS PARTICLES

DESCRIPTION

Product	RVP-768G, SARS-CoV-2 Reporter Virus Particles (RVPs)
Lot	CG-677A
Strain	BA.1 (OMICRON)
Reporter	GFP
Size	1.0 mL/vial
Packaging	20% FBS/DMEM
Viral Titer	2.35×10^6 TU/ml [†]
Recommended Input	15 μ L per well (96-well plate) for ~20% infectivity in a flow assay*
Mycoplasma Test	Negative
Expiration Date	May 2027

SAFETY & HANDLING

Shipping	Shipped on dry ice
Stability and Storage	Store at $\leq -80^\circ\text{C}$ upon receipt

* Determined in the 293T-ACE2 stable cell line

SARS-CoV-2 RVPs are used to test the ability of serum, antibodies, and drugs to neutralize the infectivity of SARS-CoV-2. RVPs display antigenically correct spike protein pseudotyped on replication-incompetent virus particles that contain a heterologous lentiviral (HIV) core. RVPs are capable of a single round of infection and carry a genome that expresses either a GFP or luciferase optical reporter gene upon infection. RVPs are produced in HEK-293T cells using three separate plasmids, encoding the spike protein, a lentiviral gag polyprotein, and a reporter gene.

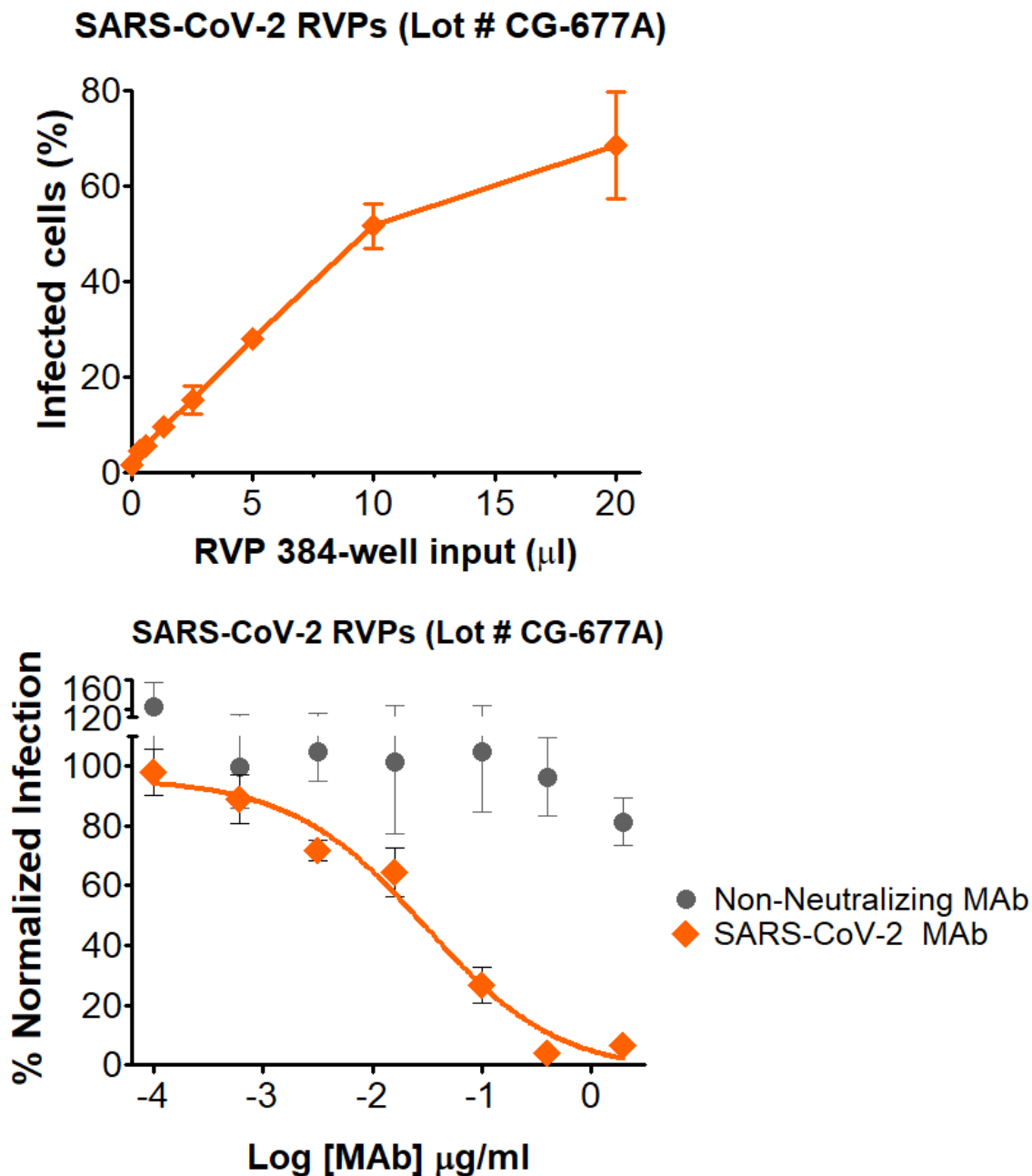
RVPs are created using a second-generation lentiviral system with components that are highly unlikely to recombine to produce a fully infectious virus (requiring 3 separate recombination events to do so). However, lentiviruses are capable of genomic integration and RVPs are derived from biological materials so should be handled with caution within a BSL2 or enhanced BSL2 laboratory environment. RVPs are not to be used in humans or in animals raised for food.

Thaw tubes in a 37°C water bath for 3 minutes and place on ice until ready to use. RVPs will appear as a translucent, pink solution. Gently mix prior to use and pulse tube for 3 seconds at high speed in a tabletop microfuge to recover all volume from the tube. Vortexing of RVPs should be avoided. Re-freezing of RVPs is not recommended.

[†] Titer is calculated using the estimated number of cells at the end of the experiment, using the following equation:

$$\left[\left(\frac{\% \text{ GFP positive cells}}{100} \right) \times \left(\frac{30,000 \text{ cells}}{1 \text{ well}} \right) \times \left(\frac{1 \text{ well}}{\text{RVP input } (\mu\text{L})} \right) \right] \times \frac{1000 \mu\text{L}}{1 \text{ mL}} = X \frac{\text{Transforming Units (TU)}}{\text{mL}}$$

INFECTIVITY & NEUTRALIZATION DATA



Infectivity and neutralization determined in HEK-293T cells stably over-expressing ACE2 (Integral Cat# C-HA102). Infectivity data represents the average of three independent vials, each tested in quadruplicate.

Neutralization utilized 5 μ l of SARS-CoV-2 RVPs in a 384-well plate. GFP positive cells were detected with an Intellicyt iQue flow cytometer using the BL-1 channel (Ex. 488 nm, Em. 530).