



## DESCRIPTION

<b>Product</b>	RVP-1401G, Zaire ebolavirus Reporter Virus Particles (RVPs)
<b>Lot</b>	FG-639B
<b>Strain</b>	Mayinga/76
<b>Reporter</b>	GFP
<b>Size</b>	1.0 mL/vial
<b>Packaging</b>	20% FBS/DMEM
<b>Viral Titer</b>	$9.50 \times 10^5$ TU/mL <sup>†</sup>
<b>Recommended Input</b>	10 $\mu$ L per well (96-well plate) for ~20% infectivity in a flow assay*
<b>Mycoplasma Test</b>	Negative
<b>Expiration Date</b>	March 2026

## SAFETY &amp; HANDLING

<b>Shipping</b>	Shipped on dry ice
<b>Stability and Storage</b>	Store at $\leq -80^\circ\text{C}$ upon receipt

\* Determined in the Huh-7 cell line

Ebola RVPs are used to test the ability of serum, antibodies, and drugs to neutralize infectivity. RVPs display antigenically correct glycoprotein pseudotyped on replication-incompetent virus particles that contain a heterologous lentiviral (HIV) core. RVPs are capable of a single round of infection and carry a genome that expresses either a GFP or luciferase optical reporter gene upon infection. RVPs are produced in HEK-293T cells using three separate plasmids, encoding the envelope glycoprotein, a lentiviral gag polyprotein, and a reporter gene.

RVPs are created using a second-generation lentiviral system with components that are highly unlikely to recombine to produce a fully infectious virus (requiring 3 separate recombination events to do so). However, lentiviruses are capable of genomic integration and RVPs are derived from biological materials so should be handled with caution within a BSL2 or enhanced BSL2 laboratory environment. RVPs are not to be used in humans or in animals raised for food.

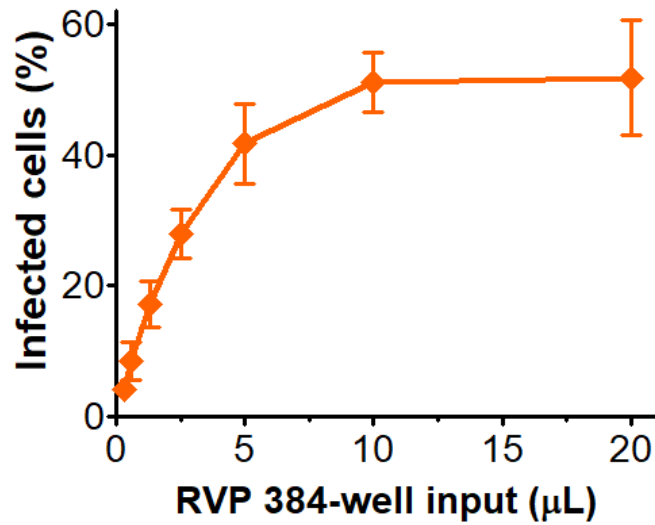
Thaw vials in a 37°C water bath for 2-3 minutes and place on ice until ready to use. RVPs will appear as a translucent, solution. Gently mix prior to use and pulse tube for 3 seconds at high speed in a tabletop microfuge to recover all volume from the tube. Vortexing of RVPs should be avoided. Re-freezing of RVPs is not recommended.

<sup>†</sup> Titer is calculated using the estimated number of cells during the time of infection, using the following equation:

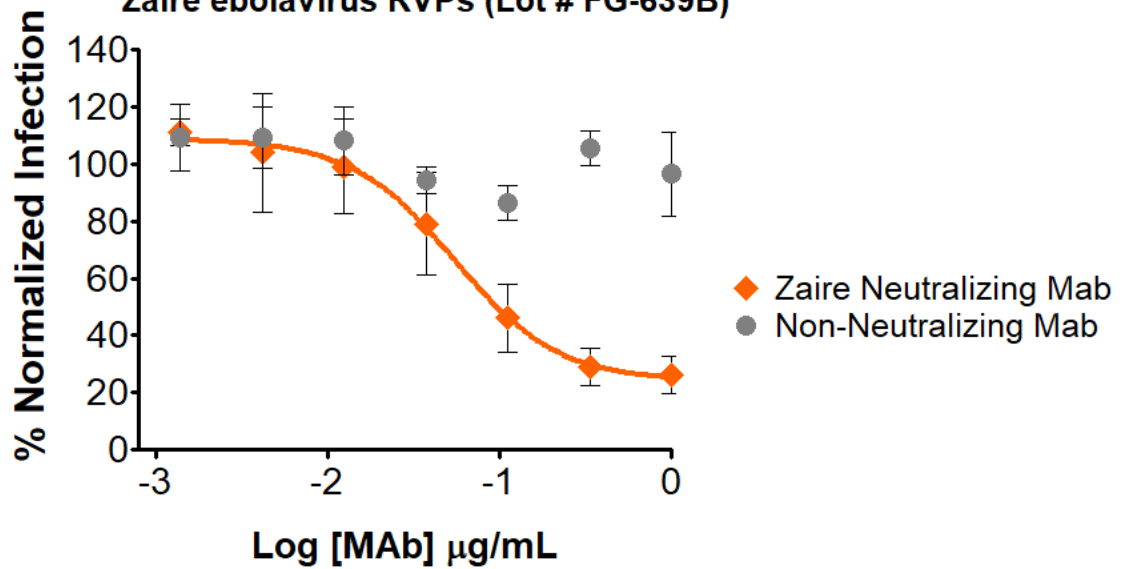
$$\left[ \left( \frac{\% \text{ GFP positive cells}}{100} \right) \times \left( \frac{9,000 \text{ cells}}{1 \text{ well}} \right) \times \left( \frac{1 \text{ well}}{\text{RVP input } (\mu\text{L})} \right) \right] \times \frac{1000 \mu\text{L}}{1 \text{ mL}} = X \frac{\text{Transforming Units (TU)}}{\text{mL}}$$

## INFECTIVITY &amp; NEUTRALIZATION DATA

## Zaire ebolavirus RVPs (Lot # FG-639B)



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Infectivity and neutralization determined in Huh-7 cells. Infectivity data represents the average of three independent vials, each tested in quadruplicate.

Neutralization utilized 10µL of Ebola RVPs in a 384-well plate. GFP positive cells were detected with an Intellicyt iQue flow cytometer using the BL-1 channel (Ex. 488 nm, Em. 530).