



# TECH DATA SHEET

## REPORTER VIRUS PARTICLES

### DESCRIPTION

<b>Product</b>	RVP-795G, SARS-CoV-2 Reporter Virus Particles (RVPs)
<b>Lot</b>	CG-598A
<b>Strain</b>	HV.1 (OMICRON)
<b>Reporter</b>	GFP
<b>Size</b>	1.0 mL/vial
<b>Packaging</b>	20% FBS/DMEM
<b>Viral Titer</b>	1.03 x 10 <sup>7</sup> TU/ml†
<b>Recommended Input</b>	2.5µL per well (96-well plate) for ~20% infectivity in a flow assay*
<b>Mycoplasma Test</b>	Negative
<b>Expiration Date</b>	November 2025

### SAFETY & HANDLING

<b>Shipping</b>	Shipped on dry ice
<b>Stability and Storage</b>	Store at ≤ -80°C upon receipt

\* Determined in the 293T-ACE2 stable cell line

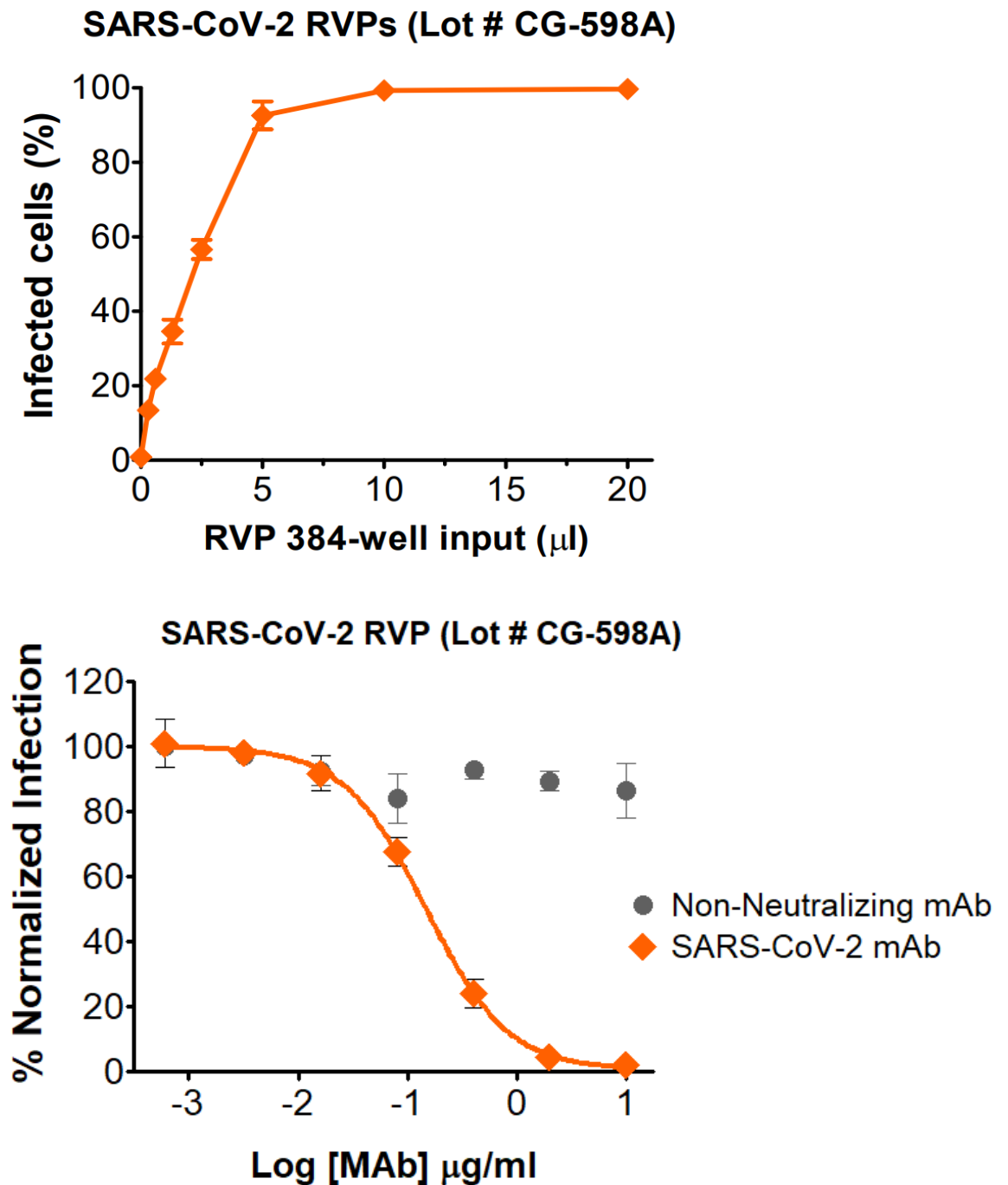
SARS-CoV-2 RVPs are used to test the ability of serum, antibodies, and drugs to neutralize the infectivity of SARS-CoV-2 spike protein. RVPs display antigenically correct spike protein pseudotyped on replication-incompetent virus particles that contain a heterologous lentiviral (HIV) core. RVPs are capable of a single round of infection and carry a genome that expresses either a GFP or luciferase optical reporter gene upon infection. RVPs are produced in HEK-293T cells using three separate plasmids, encoding the spike protein, a lentiviral gag polyprotein, and a reporter gene. RVPs are created using a second-generation lentiviral system with components that are highly unlikely to recombine to produce a fully infectious virus (requiring 3 separate recombination events to do so). However, lentiviruses are capable of genomic integration and RVPs are derived from biological materials so should be handled with caution within a BSL2 or enhanced BSL2 laboratory environment. RVPs are not to be used in humans or in animals raised for food.

Thaw tubes in a 37°C water bath for 3 minutes and place on ice until ready to use. RVPs will appear as a translucent, pink solution. Gently mix prior to use and pulse tube for 3 seconds at high speed in a tabletop microfuge to recover all volume from the tube. Vortexing of RVPs should be avoided. Re-freezing of RVPs is not recommended.

† Titer is calculated using the estimated number of cells at the end of the experiment, using the following equation:

$$\left[ \left( \frac{\% \text{ GFP positive cells}}{100} \right) \times \left( \frac{30,000 \text{ cells}}{\text{well}} \right) \times (\text{RVP input volume in } \mu\text{L}) \right] \times \frac{1000 \mu\text{L}}{1 \text{ mL}} = X \frac{\text{Transforming Units (TU)}}{\text{mL}}$$

## INFECTIVITY &amp; NEUTRALIZATION DATA



Infectivity and neutralization determined in HEK-293T cells stably over-expressing ACE2 (Integral Cat# C-HA102). Infectivity data represents the average of three independent vials, each tested in quadruplicate.

Neutralization utilized 5  $\mu\text{l}$  of SARS-CoV-2 RVPs in a 384-well plate. GFP positive cells were detected with an Intellicyt iQue flow cytometer using the BL-1 channel (Ex. 488 nm, Em. 530).