



TECH DATA SHEET

REPORTER VIRUS PARTICLES

DESCRIPTION

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|--------------------------|---|
| Product | RVP-782G, SARS-CoV-2 Reporter Virus Particles (RVPs) |
| Lot | CG-484A |
| Strain | BQ.1.1 (OMICRON) |
| Reporter | GFP |
| Size | 1.0 mL/vial |
| Packaging | 20% FBS/DMEM |
| Viral Titer | 1.01 x 10 ⁷ TU/ml |
| Recommended Input | 1.25µL per well (96-well plate) for ~20% infectivity in a flow assay* |
| Mycoplasma Test | Negative |
| Expiration Date | March 2027 |

SAFETY & HANDLING

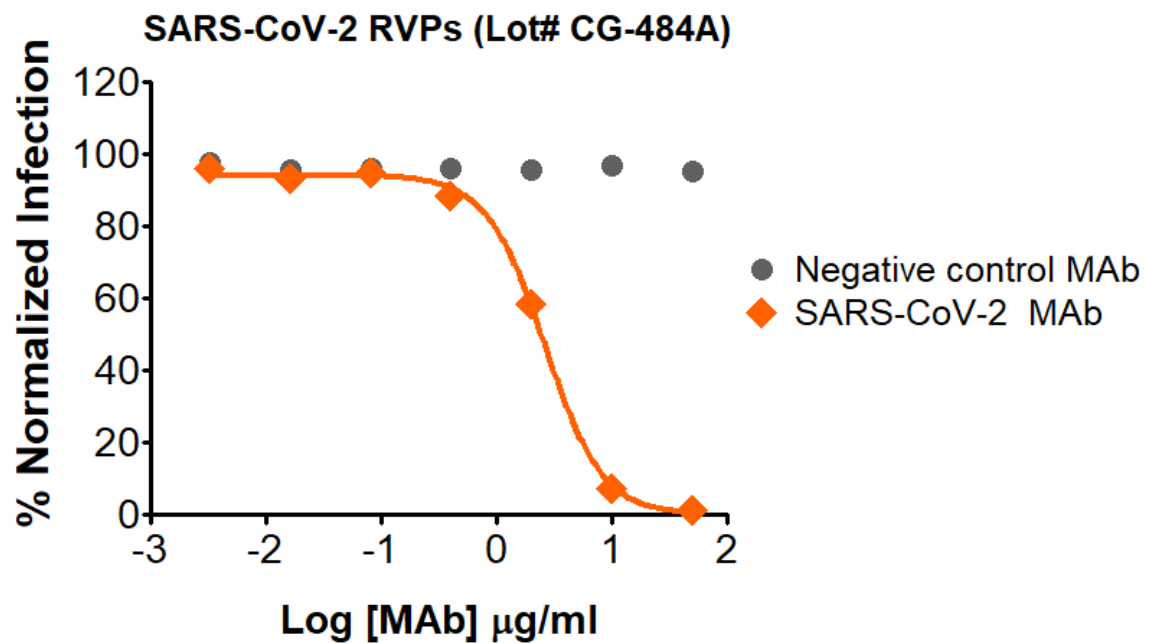
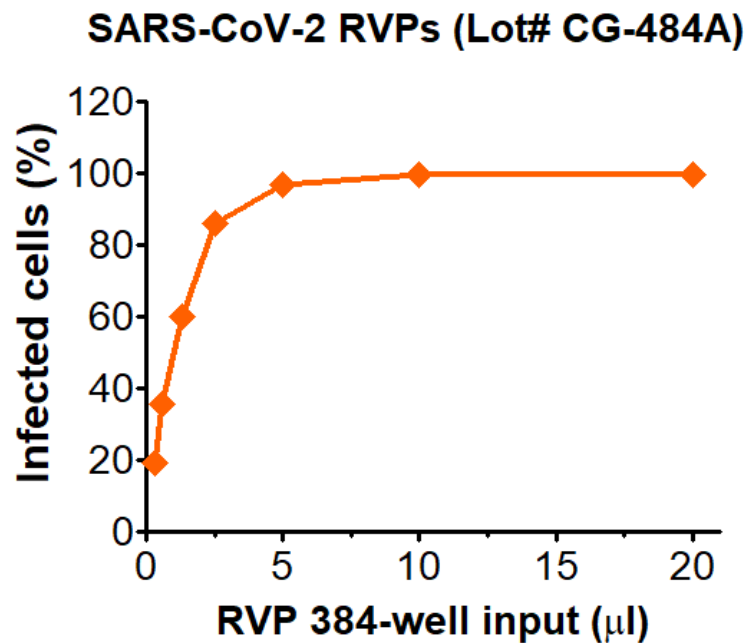
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|------------------------------|-------------------------------|
| Shipping | Shipped on dry ice |
| Stability and Storage | Store at ≤ -80°C upon receipt |

* Determined in the 293T-ACE2 stable cell line

SARS-CoV-2 RVPs are used to test the ability of serum, antibodies, and drugs to neutralize the infectivity of SARS-CoV-2 spike protein. RVPs display antigenically correct spike protein pseudotyped on replication-incompetent virus particles that contain a heterologous lentiviral (HIV) core. RVPs are capable of a single round of infection and carry a genome that expresses either a GFP or luciferase optical reporter gene upon infection. RVPs are produced in HEK-293T cells using three separate plasmids, encoding the spike protein, a lentiviral gag polyprotein, and a reporter gene. RVPs are created using a second-generation lentiviral system with components that are highly unlikely to recombine to produce a fully infectious virus (requiring 3 separate recombination events to do so). However, lentiviruses are capable of genomic integration and RVPs are derived from biological materials so should be handled with caution within a BSL2 or enhanced BSL2 laboratory environment. RVPs are not to be used in humans or in animals raised for food.

Thaw tubes in a 37°C water bath for 3 minutes and place on ice until ready to use. RVPs will appear as a translucent, pink solution. Gently mix prior to use and pulse tube for 3 seconds at high speed in a tabletop microfuge to recover all volume from the tube. Vortexing of RVPs should be avoided. Re-freezing of RVPs is not recommended.

INFECTIVITY & NEUTRALIZATION DATA



Infectivity and neutralization determined in HEK-293T cells stably over-expressing ACE2 (Integral Cat# C-HA102). Infectivity data represents the average of three independent vials, each tested in quadruplicate.

Neutralization utilized 5 µl of SARS-CoV-2 RVPs in a 384-well plate. GFP positive cells were detected with an Intellicyt iQue flow cytometer using the BL-1 channel (Ex. 488 nm, Em. 530).